

# Detection of Desmin in Formalin-Fixed, Paraffin Embedded Rat Tissue

## Reagents:

[1X Automation Buffer](#)

[3% Hydrogen Peroxide](#)

[Antibody Diluent](#)

[DAB Chromagen](#)

[Hematoxylin](#)

## Antibody Information:

Kit : Vector Rabbit Elite Kit

Vector Laboratories, Inc.

Burlingame, CA 94010

[www.vectorlabs.com](http://www.vectorlabs.com)

1-800-227-6666

Catalog: PK-6101

\*The Vector Rabbit Elite Kit contains solutions needed to make the secondary and label antibodies.

Blocking Serum: Normal Goat Serum

Jackson ImmunoResearch Laboratories, Inc.

West Grove, PA 19390

[www.jacksonimmuno.com](http://www.jacksonimmuno.com)

1-800-367-5296

Catalog #005-000-001

Avidin Biotin Blocking Kit

Vector Laboratories, Inc.

Burlingame, CA 94010

[www.vectorlabs.com](http://www.vectorlabs.com)

1-800-227-6666

Catalog #SP-2001

Primary Antibody: Rabbit anti-desmin

Accurate Chemical & Scientific Corp.

Westbury, NY 11590

(516) 333-2221

[www.accuratechem.com](http://www.accuratechem.com)

Catalog #YMPS31

Negative control serum: Normal Rabbit Serum  
Jackson ImmunoResearch Laboratories, Inc.  
West Grove, PA 19390  
[www.jacksonimmuno.com](http://www.jacksonimmuno.com)  
1-800-367-5296  
Catalog #011-000-001

### Staining Procedure

Positive Control Tissue: Smooth and striated muscle cells of the heart  
Stain localization: Cytoplasmic

Deparaffinize and hydrate slides through the following solutions.

Xylene	2 times	5 minutes
100% Ethanol	2 times	3 minutes
95% Ethanol	2 times	3 minutes
1X Automation Buffer	2 times	5 minutes

1. Quench endogenous peroxidase by placing slides in 3% hydrogen peroxide for 15 minutes.
2. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
3. Unmasking Techniques using the microwave oven.  
Place a full rack of slides in tissue tekô container containing 200ml of distilled water.  
Microwave for 5 minutes at level 5  
Cool for 1 minute (Add more distilled water if necessary)  
Microwave for 5 minutes at level 5. Temp after Microwaving \_\_\_\_\_  
Cool 20 minutes at room temperature.
4. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.
5. Block with 5% Normal Goat Serum for 20 minutes at room temperature.  
Lot # \_\_\_\_\_ Reconstituted Date \_\_\_\_\_
6. Apply Avidin/Biotin block  
Lot# \_\_\_\_\_ Exp. Date \_\_\_\_\_ New Kit:    yes /    no  
Apply avidin block - 15 minutes at room temperature.  
Quick rinse in 1X AB.  
Apply biotin block - 15 minutes at room temperature.  
Wipe excess block.

7. Apply primary antibody (Desmin) at 1:500 dilution and incubate for one hour at room temperature.

Lot # \_\_\_\_\_ Exp date \_\_\_\_\_

For negative control slides, normalize the protein concentration of the normal rabbit serum with the protein concentration of the primary antibody (Desmin) and use this to make the 1:500 dilution. Apply to slides and incubate for one hour at room temperature.

Lot# \_\_\_\_\_ Reconstituted Date \_\_\_\_\_

8. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.

9. Apply secondary antibody and incubate for 30 minutes at room temperature.

Exp date \_\_\_\_\_ New Kit:    yes    /    no

10. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.

11. Apply Label antibody and incubate for 30 minutes at room temperature.  
(Prepare at least 30 minutes prior to use)

12. Rinse slides in 2 changes of 1X Automation Buffer for 5 minutes each.

13. Apply liquid Dako DAB Chromagen for 6 minutes in the dark.  
(Add 1 drop of DAB per ml of substrate)

Lot# \_\_\_\_\_ Exp. Date \_\_\_\_\_ New Kit:    yes    /    no

14. Rinse in tap water 3 minutes.

15. Counterstain with Modified Harris Hematoxylin for 20 seconds.

16. Rinse in tap water until water is clear.

17. Gently agitate slides in 1X Automation Buffer until blue.

18. Dehydrate through the following solutions.

95% Ethanol	1 change	3 minutes
100% Ethanol	3 changes	3 minutes
Xylene	2 changes	5 minutes

19. Coverslip

*Updated 08/24/06*